

Available online at www.sciencedirect.com



CARBOHYDRATE RESEARCH

Carbohydrate Research 338 (2003) 843-849

www.elsevier.com/locate/carres

Regioselective glycosylation of 4,6-O-benzylidenated glucopyranosides

Ying Zeng, Fanzuo Kong*

Research Center for Eco-Environmental Sciences, Academia Sinica, Chinese Academy of Sciences, P.O. Box 2871, Beijing 100085, China Received 22 November 2002; accepted 13 January 2003

Abstract

Regioselective glycosylation with allyl 4,6-O-benzylidene- α , β -D-glucopyranoside or methyl 4,6-O-benzylidene- α , β -D-glucopyranoside as the acceptor was investigated. It was found that the regioselectivity depends upon donor size and anomeric configuration of the acceptor, i.e., with a monosaccharide donor and an α -form acceptor, the $(1 \rightarrow 3)$ -linked product was obtained predominantly or exclusively, while with disaccharide or trisaccharide donors and either an α or β form acceptor, the $(1 \rightarrow 2)$ -linked oligosaccharides were the only products. © 2003 Elsevier Science Ltd. All rights reserved.

Keywords: Glucose oligosaccharides; Trichloroacetimidates; Regioselective glycosylation

1. Introduction

Regioselective coupling is an effective and concise method for the synthesis of oligosaccharides as it can simplify the synthetic route substantially and avoid a tedious protection—deprotection procedure. Just like stereoselective formation of the glycosidic linkage, the results of regioselective coupling are influenced by many factors. Recently, a series of work on regioselective coupling has been carried out in our group, resulting in a variety of branched mannose, rhamnose, and glucose oligosaccharides.²

β-(1 \rightarrow 3)-Linked glucans occur in some biologically important natural products, such as schizophyllan, scleroglucan and lentinan with antitumor activity,³ and the β-(1 \rightarrow 2)-glucosyl linkage occurs in the capsular polysaccharide of *Streptococcus peumoniae* type 37^{4a} and in the exopolysaccharide of *Pediococus domnosus*.^{4b} 4,6-*O*-Benzylidenated glucopyranosides are good acceptors since their 3-*O*- or 2-*O*-selective coupling with appropriate donors can afford (1 \rightarrow 3)- or (1 \rightarrow 2)-linked oligosaccharides, based on which useful fragments of natural polysaccharides can be synthesized. However,

E-mail address: fzkong@mail.rcees.ac.cn (F. Kong).

there is a very subtle distinction between the 2-OH and the 3-OH of the acceptors, and the activity of the 3-OH is little bit greater than that of the 2-OH when fully benzylated glucosyl trichloroacetimidate was used as the donor.⁵ We present herein 3-*O*- or 2-*O*-regioselective glycosylation of 4,6-*O*-benzylidenated glucosides with fully benzoylated glycosyl trichloroacetimidates as donors.

2. Results and discussion

As shown in Scheme 1, glycosylation of methyl 4,6-Obenzylidene-α-D-glucopyranoside (2) with 2,3,4,6-tetra-O-benzoyl- α -D-glucopyranosyl trichloroacetimidate (1) afforded a mixture of β -(1 \rightarrow 3)-linked disaccharide 3 (53%) and β -(1 \rightarrow 2)-linked disaccharide 4 (24%), and no α-linked product was detected. Compounds 3 and 4 were isolated and characterized by their acetylation to give the corresponding 2-acetate and 3-acetate, respectively. The former showed H-2 at δ 4.77 ppm with $J_{1,2} = 4.0$, $J_{2,3} = 9.6$ Hz, and the latter showed H-3 at δ 5.46 ppm with $J_{2,3} = J_{3,4} = 9.6$ Hz in their ¹H NMR spectra. It was interesting to find that coupling of 1 with the allyl glucoside 5 instead of 2, gave exclusive β -(1 \rightarrow 3)-linked disaccharide 6 (75%), disclosing that allyl, instead of methyl, as the aglycon moiety of the acceptor led to 3-O-glycosylation. It was also found

^{*} Corresponding author. Tel.: + 86-10-62936613; fax: + 86-10-62923563

that condensation of 5 with the donor, 2,3-di-O-acetyl-4,6-O-benzylidene-α-D-glucopyranosyl trichloroacetimidate (7), yielded solely the β -(1 \rightarrow 3)-linked disaccharide 8 (81%). To examine the effect of anomeric configuration in the acceptor, methyl 4,6-O-benzylidene-β-Dglucopyranoside (9) and allyl 4,6-O-benzylidene-β-Dglucopyranoside (12) were used as the acceptors for coupling with 1. Again, the regioselectivity was established by acetylation. In the ¹H NMR spectra of the acetylated compounds, H-2 was found by irradiation of H-1. For the 3-O-glycosylation, H-2 appeared at δ ~ 5.0 ppm, whereas for the 2-O-glycosylation, H-2 appeared at δ 3.58–3.83 ppm. It was found that the regioselectivity was changed substantially along with the configuration alteration, i.e., β -glucoside 9 inversely gave $(1 \rightarrow 2)$ -linked 11 (63%) as the major product and

 $(1 \rightarrow 3)$ -linked **10** (20%) as the minor one. Meanwhile, β -glucoside **12** afforded $(1 \rightarrow 2)$ -linked **14** and $(1 \rightarrow 3)$ -linked **13** in almost equivalent amounts. However, glycosylation of the acceptor **2** or **5** or their β anomer **9** or **12** with β - $(1 \rightarrow 3)$ -linked disaccharide **17** or β - $(1 \rightarrow 6)$ -linked disaccharide **15** or 3,6-branched trisaccharide **22**, always furnished $(1 \rightarrow 2)$ -linked oligosaccharides, indicating that large size donors intended to give $(1 \rightarrow 2)$ -regioselectivity (Scheme 2).

Vasella's group⁵ explored the regioselectivity–structure relationship. They found that the glycosylation regioselectivity depends upon the relative strength of the intramolecular H-bonds of the acceptors when the donors were monosaccharides. Fraser-Reid and coworkers⁶ found that a 2-*O*-benzoyl group in the donor gave more 2-linked product compared to the 2-*O*-ben-

Scheme 1.

Scheme 2.

zyl group when the acceptors were methyl 4,6-O-benzylidene- α , β -D-glucopyranosides. In our research, di- or trisaccharide donors exclusively gave 2-O-glycosylation products, and the β -form of the acceptor tended to give 2-O-glycosylation. We hypothesized that the large-size donors had a serious repulsion with the benzylidene group of the acceptor, making 3-O less reactive than 2-O, and the β -form of the acceptor had less steric hindrance between the donor and aglycon, leading to more of the $(1 \rightarrow 2)$ -linkage.

In summary, the regioselectivity of glycosylation of a 4,6-O-benzylidene glucopyranoside depends upon donor size and anomeric configuration of acceptor.

With a monosaccharide donor and an α -form acceptor, $(1 \rightarrow 3)$ -linked product was obtained predominantly or exclusively, while with di- or trisaccharide as the donors, $(1 \rightarrow 2)$ -linked oligosaccharides were the only products.

3. Experimental

3.1. General methods

Optical rotations were determined at 25 °C with a Perkin–Elmer Model 241-Mc automatic polarimeter.

 1 H, 13 C NMR were recorded with Bruker ARX 400 spectrometers (400 MHz for 1 H, 100 MHz for 13 C) at 25 °C for solutions in CDCl₃ or D₂O as indicated. Thin-layer chromatography (TLC) was performed on silica gel HF₂₅₄ with detection by charring with 30% (v/v) H₂SO₄ in MeOH or in some cases by a UV lamp. Column chromatography was conducted by elution of a column (8 × 240, 18 × 300, 35 × 400 mm) of silica gel (100–200 mesh) with EtOAc–petroleum ether (bp 60–90 °C) as the eluent. Solutions were concentrated at < 60 °C under reduced pressure.

3.2. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -4,6-O-benzylidene- α -D-glucopyranoside (3)

Compounds 2 (150 mg, 0.20 mmol) and 1 (60 mg, 0.21 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd CH₂Cl₂ (5 mL). TMSOTf (8 μ L) was added dropwise at -20 °C with N_2 protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually raised to ambient temperature. Then the mixture was neutralized with Et₃N. Concentration of the reaction mixture, followed by purification on a silica gel column with 2:1 petroleum ether-EtOAc as the eluent, gave two disaccharides with R_f 0.45 and 0.60 on TLC (2:1 petroleum ether-EtOAc), respectively. The disaccharide with R_f 0.45 was acetylated in pyridine (2 mL) with Ac₂O (0.5 mL) for 3 h, then concentrated to dryness. The residue was purified by flash chromatography (3:1 petroleum ether-EtOAc) to give the 2-acetate of 3 (95 mg, 53% for two steps) as a syrup: $[\alpha]_D + 17.0^\circ$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.93–7.25 (m, 25 H, Ph*H*), 5.85 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.65 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 9.6$ Hz, H-3'), 5.600 (s, 1 H, PhCH), 5.50 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 5.07 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 4.85 (d, 1 H, $J_{1,2}$ 4.0 Hz, H-1), 4.77 (dd, 1 H, $J_{2,3}$ 9.6, $J_{1,2}$ 4.0 Hz, H-2), 4.47 (dd, 1 H, $J_{6'a,6'b}$ 12.0, $J_{5',6'a}$ 3.6 Hz, H-6'a), 4.30–4.24 (m, 3 H), 3.91-3.73 (m, 4 H), 3.32 (s, 3 H, CH_3O), 1.77 (s, 3 H, CH_3CO). Anal. Calcd for $C_{50}H_{46}O_{16}$: C, 66.51; H, 5.14. Found: C, 66.79; H, 5.01.

3.3. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- α -D-glucopyranoside (4)

The disaccharide with R_f 0.60 in Section 3.2, above, was acetylated with Ac₂O in pyridine to give the 3-acetate of 4 (45 mg, 24% for two steps): $[\alpha]_D + 13.2^\circ$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.03–7.25 (m, 25 H, PhH), 5.93 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.69 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 9.6$ Hz, H-3'), 5.55 (dd, 1 H, $J_{1',2'}$ 7.8, $J_{2',3'}$ 9.6 Hz, H-2'), 5.46 (dd, 1 H, $J_{3,4} = J_{2,3} = 9.6$ Hz, H-3), 5.41 (s, 1 H, PhCH), 5.01 (d, 1 H, $J_{1,2}$ 3.2 Hz, H-1), 4.99 (d, 1 H, $J_{1',2'}$ 7.8 Hz, H-1'), 4.77–3.85 (m, 5 H), 3.74 (dd, $J_{1,2}$ 3.2, $J_{2,3}$ 9.6 Hz, H-2),

3.71–3.46 (m, 2 H), 3.37 (s, 3 H, CH_3O), 1.70 (s, CH_3CO). Anal. Calcd for $C_{50}H_{46}O_{16}$: C, 66.51; H, 5.14. Found: C, 66.27; H, 5.22.

3.4. Allyl 2,3,4,6-tetra-*O*-benzoyl-β-D-glucopyranosyl-(1 → 3)-4,6-*O*-benzylidene-α-D-glucopyranoside (6)

The 2-acetate of 6 (138 mg, 75% for two steps) was obtained as a syrup by coupling of 1 (150 mg, 0.20 mmol) with 5 (62 mg, 0.20 mmol), followed by acetylation under the same conditions as described for the preparation of the 2-acetate of 3: $[\alpha]_D + 15.3^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.95–7.26 (m, 25 H, PhH), 5.82 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.81-5.72 (m, 1 H, -CH=), 5.66 (dd, 1 H, $J_{3',4'}=J_{2',3'}=$ 9.6 Hz, H-3'), 5.60 (s, 1 H, PhCH), 5.50 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 5.22–5.13 (m, 2 H, =C H_2), 5.09 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 5.00 (d, 1 H, $J_{1,2}$ 3.6 Hz, H-1), 4.76 (dd, 1 H, $J_{1.2}$ 4.0, $J_{2.3}$ 9.6 Hz, H-2), 4.47 (dd, 1 H, $J_{6'a,6'b}$ 12.4, $J_{5',6'a}$ 3.2 Hz, H-6'a), 4.34–4.23 (m, 3 H), 4.10-3.89 (m, 4 H), 3.79-3.74 (m, 2 H), 1.74 (s, 3 H, CH_3CO). Anal. Calcd for $C_{52}H_{48}O_{16}$: C, 67.27; H, 5.21. Found: C, 67.43; H, 5.19.

3.5. Allyl 2,3-di-O-acetyl-4,6-O-benzylidene- β -D-glucopyranosyl-(1 \rightarrow 3)-4,6-O-benzylidene- α -D-glucopyranoside (8)

The 2-acetate of 8 (110 mg, 81% for two steps) was obtained as a syrup by coupling of 7 (100 mg, 0.20) mmol) with 5 (62 mg, 0.20 mmol), followed by acetylation under the same conditions as described for the preparation of the 2-acetate of 3: $[\alpha]_D + 15.3^\circ$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.51–7.34 (m, 10 H, PhH), 5.91-5.82 (m, 1 H, -CH=), 5.57 (s, 1 H, PhCH), 5.34 (s, 1 H, PhCH), 5.33-5.21 (m, 2 H, $=CH_2$), 5.21 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 9.6$ Hz, H-3'), 5.05 (d, 1 H, $J_{1,2}$ 4.0 Hz, H-1), 5.01 (dd, 1 H, $J_{1',2'}$ 7.8, $J_{2',3'}$ 9.6 Hz, H-2'), 4.88 (d, 1 H, $J_{1',2'}$ 7.8 Hz, H-1'), 4.86 (dd, $J_{1,2}$ 4.0, $J_{2,3}$ 9.6 Hz, H-2), 4.32–4.16 (m, 4 H), 4.02– 3.89 (m, 2 H), 3.79–3.63 (m, 4 H), 3.49–3.42 (m, 1 H), 2.16 (s, 3 H, CH₃CO), 2.04 (s, 6 H, 2CH₃CO). Anal. Calcd for C₃₅H₄₀O₁₄: C, 61.40; H, 5.89. Found: C, 61.31; H, 5.97.

3.6. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -4,6-O-benzylidene- β -D-glucopyranoside (10)

The 2-acetate of **10** (115 mg, 63% for two steps) was obtained as a syrup by coupling of **1** (150 mg, 0.20 mmol) with **9** (60 mg, 0.21 mmol), followed by silica gel separation and acetylation under the same conditions as described for the preparation of the 2-acetate of **3**: $[\alpha]_D + 13.5^\circ$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.92–7.26 (m, 25 H, Ph*H*), 5.79 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.63 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 3.6$

9.6 Hz, H-3'), 5.58 (s, 1 H, PhC*H*), 5.46 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 5.05 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 4.98 (dd, 1 H, $J_{1,2}$ 8.0, $J_{2,3}$ 8.8 Hz, H-2), 4.45–4.33 (m, 2 H), 4.30 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 4.24–4.20 (m, 1 H), 4.02–3.97 (m, 1 H), 3.84–3.77 (m, 3 H), 3.47–3.42 (m, 1 H), 3.37 (s, 3 H, CH_3O), 1.73 (s, 3 H, CH_3CO). Anal. Calcd for $C_{50}H_{46}O_{16}$: C, 66.51; H, 5.14. Found: C, 66.32; H, 5.17.

3.7. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- β -D-glucopyranoside (11)

The 3-acetate of 11 (35 mg, 20%, two steps) was prepared as a syrup under the same conditions as described for the preparation of the 3-acetate of 4: $[\alpha]_D + 16.8^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.92–7.24 (m, 25 H, PhH), 5.85 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6 \text{ Hz}, \text{ H-4'}, 5.72 \text{ (dd, 1 H, } J_{3',4'} = J_{2',3'} =$ 9.6 Hz, H-3'), 5.48 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 5.37 (s, 1 H, PhCH), 5.25 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 5.19 (dd, 1 H, $J_{3,4} = J_{2,3} = 9.6$ Hz, H-3), 4.67 (dd, 1 H, $J_{6'a,6'b}$ 12.4, $J_{5',6'a}$ 3.2 Hz, H-6'a), 4.57 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 4.48 (dd, 1 H, $J_{6'a,6'b}$ 12.4, $J_{5',6'b}$ 3.2 Hz, H-6'b), 4.28 (dd, 1 H, $J_{6a,6b}$ 12.0, $J_{5,6a}$ 3.2 Hz, H-6a), 4.18 (dd, 1 H, $J_{6a,6b}$ 12.0, $J_{5,6b}$ 3.2 Hz, H-6b), 3.76 (dd, 1 H, $J_{1,2}$ 8.0, $J_{2,3}$ 9.6 Hz, H-2), 3.69–3.55 (m, 2 H), 3.51 (s, 3 H, CH_3O), 3.79–3.45 (m, 1 H), 1.73 (s, 3 H, CH_3CO). Anal. Calcd for $C_{50}H_{46}O_{16}$: C, 66.51; H, 5.14. Found: C, 66.61; H, 5.20.

3.8. Allyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -4,6-O-benzylidene- β -D-glucopyranoside (13)

The 2-acetate of 13 (77 mg, 41% for two steps) was obtained as a syrup by coupling of 1 (150 mg, 0.20 mmol) with 12 (62 mg, 0.20 mmol), followed by silica gel separation and acetylation under the same conditions as described for the preparation of the 2-acetate of 3: $[\alpha]_D + 8.0^{\circ}$ (c 2.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.95–7.25 (m, 25 H, PhH), 5.81 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6 \text{ Hz}, \text{ H-4'}, 5.76-5.70 (m, 1 \text{ H}, -\text{C}H=),$ 5.64 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 9.6$ Hz, H-3'), 5.59 (s, 1 H, PhCH), 5.48 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 5.20-5.09 (m, 2 H, = CH_2), 5.05 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 5.03 (dd, 1 H, $J_{1,2}$ 8.0, $J_{2,3}$ 8.8 Hz, H-2), 4.47– 4.43 (m, 1 H), 4.43 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 4.33 (m, 1 H), 4.26–4.22 (m, 2 H), 4.04–3.95 (m, 2 H), 3.91– 3.78 (m, 3 H), 3.47-3.39 (m, 1 H), 1.73 (s, 3 H, CH_3CO). Anal. Calcd for $C_{52}H_{48}O_{16}$: C, 67.27; H, 5.21. Found: C, 66.97; H, 5.20.

3.9. Allyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- β -D-glucopyranoside (14)

The 3-acetate of 14 (75 mg, 41%, two steps) was prepared as a syrup under the same conditions as

described for the preparation of the 3-acetate of 4: $[\alpha]_D + 6.5^\circ$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.04–7.25 (m, 25 H, PhH), 5.95–5.84 (m, 1 H, CH=), 5.86 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.75 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 9.6$ Hz, H-3'), 5.49 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 5.38 (s, 1 H, PhCH), 5.36–5.31 (m, 1 H, =C H_{2a}), 5.28 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 5.21 (dd, 1 H, $J_{3,4} = J_{2,3} = 9.2$ Hz, H-3), 5.19–5.15 (m, 1 H, =C H_{2b}), 4.72 (d, 1 H, $J_{1,2}$ 7.8 Hz, H-1), 4.64 (dd, 1 H, $J_{6'a,6'b}$ 12.4, $J_{5',6'a}$ 3.2 Hz, H-6'a), 4.47–4.13 (m, 5 H), 3.83 (dd, 1 H, $J_{1,2}$ 8.0, $J_{2,3}$ 9.2 Hz, H-2), 3.68–3.46 (m, 3 H), 1.76 (s, 3 H, C H_3 CO). Anal. Calcd for $C_{52}H_{48}O_{16}$: C, 67.27; H, 5.21. Found: C, 67.37; H, 5.31.

3.10. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 6)$ -2,4-di-O-acetyl-3-O-allyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- α -D-glucopyranoside (16)

2,3,4,6-Tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 6)$ -2,4-di-O-acetyl-3-O-allyl-α-D-glucopyranosyl trichloroacetimidate (15, 100 mg, 0.1 mmol) and methyl 4,6-Obenzylidene-α-D-glucopyranoside (2, 30 mg, 0.11 mmol) were dried together under high vacuum for 2 h, then dissolved in anhyd CH₂Cl₂ (5 mL). TMSOTf (8 μL) was added dropwise at -20 °C with N_2 protection. The reaction mixture was stirred for 3 h, during which time the temperature was gradually raised to ambient temperature. Then the mixture was neutralized with Et₃N. Concentration of the reaction mixture, followed by purification on a silica gel column with 2:1 petroleum ether-EtOAc as the eluent, gave the trisaccharide (72 mg, 63%). The trisaccharide was acetylated in pyridine (2 mL) with Ac₂O (0.5 mL), then concentrated to dryness. The residue was purified by flash chromatography (2:1 petroleum ether-EtOAc) to give the 3-acetate of 16 (70 mg, 94%) as a syrup: $[\alpha]_D + 12.0^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.06–7.25 (m, 25 H, PhH), 5.88 (dd, 1 H, $J_{3'',4''} = J_{4'',5''} = 9.6$ Hz, H-4"), 5.72-5.68 (m, 1 H, -CH=), 5.68 (dd, 1 H, $J_{3'',4''}=J_{2'',3''}=9.6$ Hz, H-3"), 5.57–5.47 (m, 3 H, H-2", H-3, PhCH), 5.22–5.15 (m, 2 H, CH_2 =), 5.01 (d, 1 H, $J_{1'',2''}$ 8.0 Hz, H-1"), 4.86 (dd, 1 H, $J_{4',5'} = J_{4',3'}$ 9.6 Hz, H-4'), 4.80 (d, 1 H, $J_{1,2}$ 4.0 Hz, H-1), 4.78 (dd, 1 H, $J_{2',3'}$ 9.6, $J_{1',2'}$ 8.0 Hz, H-2'), 4.65-4.48 (m, 2 H), 4.45 (dd, 1 H $J_{1',2'}$ 8.0 Hz, H-1'), 4.30-4.12 (m, 2 H), 3.96-3.75 (m, 4 H), 3.71 (dd, 1 H, $J_{2.3}$ 9.6, $J_{1.2}$ 4.0 Hz, H-2), 3.67 (dd,1 H, $J_{3.4} = J_{4.5} = 9.6$ Hz, H-4), 3.65-3.63 (m, 1 H), 3.43 (dd, 1 H, $J_{3',4'}$ $J_{2',3'} = 9.6 \text{ Hz}, \text{ H-3'}$, 3.36 (s, 3 H, C H_3 O), 2.09 (s, 3 H, CH_3CO), 2.07 (s, 3 H, CH_3CO), 1.99 (s, 3 H, CH_3CO). Anal. Calcd for C₆₃H₆₄O₂₃: C, 63.63; H, 5.42. Found: C, 64.01; H, 5.50.

3.11. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -2,4,6-tri-O-acetyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- α -D-glucopyranoside (18)

The 3-acetate of 18 (111 mg, 64% for two steps) was obtained as a syrup by coupling of 2,3,4,6-tetra-O-benzoyl-β-D-glucopyranosyl- $(1 \rightarrow 3)$ -2,4,6-tri-O-acetyl- α -Dglucopyranosyl trichloroacetimidate (17, 150 mg, 0.15 mmol) with 2 (40 mg, 0.15 mmol) followed by acetylation under the same conditions as described for the preparation of the 3-acetate of 16: $[\alpha]_D + 21.4^\circ$ (c 0.5, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.03–7.26 (m, 25 H, PhH), 5.90 (dd, 1 H, $J_{3'',4''} = J_{4'',5''} = 9.6$ Hz, H-4"), 5.68 (dd, 1 H, $J_{3'',4''} = J_{2'',3''} = 9.6$ Hz, H-3"), 5.43 (s, 1 H, PhCH), 5.43-5.38 (m, 2 H, H-4', H-2"), 5.03 (dd, 1 H, $J_{3,4} = J_{2,3} = 9.6$ Hz, H-3), 4.92 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 4.89 (d, 1 H, $J_{1'',2''}$ 8.0 Hz, H-1"), 4.74 (d, 1 H, J_{1,2} 3.6 Hz, H-1), 4.61–4.50 (m, 2 H), 4.47 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 4.38–4.07 (m, 4 H), 3.97-3.69 (m, 3 H), 3.62 (dd, 1 H, $J_{1.2}$ 4.0, $J_{2.3}$ 9.6 Hz, H-2), 3.54-3.46 (m, 2 H), 3.32 (s, 3 H, CH_3O), 2.08, 2.06, 1.97, 1.90 (4 s, 12 H, 4 CH₃CO). Anal. Calcd for $C_{62}H_{62}O_{24}$: C, 62.52; H, 5.25. Found: C, 62.27; H, 5.31.

3.12. Allyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -2,4,6-tri-O-acetyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- α -D-glucopyranoside (19)

The 3-acetate of 19 (105 mg, 59% for two steps) was obtained as a syrup by coupling of 17 (150 mg, 0.15 mmol) with 5 (45 mg, 0.15 mmol), followed by acetylation under the same conditions as described for the preparation of the 3-acetate of 16: $[\alpha]_D + 8.2^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.04–7.29 (m, 25 H, Ph*H*), 5.92 (dd, 1 H, $J_{3'',4''} = J_{4'',5''} = 9.6$ Hz, H-4"), 5.85 (m, 1 H, -CH=), 5.70 (dd, 1 H, $J_{3'',4''}=$ $J_{2'',3''} = 9.6 \text{ Hz}, \text{ H-3''}, 544 \text{ (s, 1 H, PhC}H), 5.43 \text{ (dd, 1)}$ H, $J_{1'',2''}$ 8.0, $J_{2'',3''}$ 9.6 Hz, H-2"), 5.32-5.15 (m, 2 H, CH_2 =), 5.04 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.02 (d, 1 H, $J_{1'',2''}$ 4.0 Hz, H-1"), 4.95–4.90 (m, 3 H, H-2', H-3, H-1), 4.65-4.54 (m, 2 H), 4.50 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1'), 4.30–3.95 (m, 7 H), 3.72–3.69 (m, 1 H), 3.66 (dd, 1 H, $J_{1,2}$ 4.0, $J_{2,3} = 9.6$ Hz, H-2), 3.59–3.50 (m, 2 H), 2.09, 2.09, 1.99, 1.91 (3 s, 12 H, 4 CH₃CO). Anal. Calcd for C₆₄H₆₄O₂₄: C, 63.15; H, 5.30. Found: C, 63.35; H, 5.30.

3.13. Methyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -2,4,6-tri-O-acetyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- β -D-glucopyranoside (20)

The 3-acetate of **20** (121 mg, 70% for two steps) was prepared as a syrup by coupling of **17** (150 mg, 0.15 mmol) with **9** (40 mg, 0.15 mmol), followed by acetylation under the same conditions as described for the preparation of the 3-acetate of **16**: $[\alpha]_D - 15.3^\circ$ (*c* 1.0,

CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.02–7.26 (m, 25 H, Ph*H*), 5.90 (dd, 1 H, $J_{3'',4''} = J_{4'',5''} = 9.6$ Hz, H-4"), 5.65 (dd, 1 H, $J_{3'',4''} = J_{2'',3''} = 9.6$ Hz, H-3"), 5.44 (s, 1 H, PhC*H*), 5.42 (dd, 1 H, $J_{1'',2''}$ 8.0, $J_{2'',3''}$ 9.6 Hz, H-2"), 5.09 (dd, 1 H, $J_{3',4'} = J_{4',5'} = 9.6$ Hz, H-4'), 5.00 (dd, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.6 Hz, H-2'), 4.98 (d, 1 H, $J_{1'',2''}$ 8.0 Hz, H-1"), 4.89 (dd, 1 H, $J_{3,4} = J_{2,3} = 9.6$ Hz, H-3), 4.70 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 4.67–4.50 (m, 2 H), 4.34 (d, 1 H, $J_{1,2}$ 3.2 Hz, H-1), 4.33–4.29 (m, 1 H), 4.28–4.11 (m, 2 H), 3.93–3.91 (m, 1 H), 3.68–3.66 (m, 1 H), 3.64 (dd, $J_{1,2}$ 8.0, $J_{2,3}$ 9.6 Hz, H-2), 3.64–3.62 (m, 1 H), 3.59–3.54 (m, 1 H), 3.47 (s, 3 H, C*H*₃O), 3.41–3.34 (m, 1 H), 2.06, 2.04, 1.96, 1.95 (4 s, 12 H, 4 C*H*₃CO). Anal. Calcd for $C_{62}H_{62}O_{24}$: C, 62.52; H, 5.25. Found: C, 62.48; H, 5.41.

3.14. Allyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -2,4,6-tri-O-acetyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- β -D-glucopyranoside (21)

The 3-acetate of 21 (105 mg, 59% for two steps) was obtained as a syrup by coupling of 17 (150 mg, 0.15 mmol) with 12 (45 mg, 0.15 mmol), followed by acetylation under the same conditions as described for the preparation of the 3-acetate of 16: $[\alpha]_D - 25.5^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.95–7.18 (m, 25 H, Ph*H*), 5.82 (dd, 1 H, $J_{3'',4''} = J_{4'',5''} = 9.6$ Hz, H-4"), 5.80 (m, 1 H, -CH=), 5.60 (dd, 1 H, $J_{3'',4''}=$ $J_{2'',3''} = 9.6 \text{ Hz}, \text{ H-3''}, 5.36 \text{ (s, 1 H, PhC}H), 5.34 \text{ (dd, 1)}$ H, $J_{1'',2''}$ 8.0, $J_{2'',3''}$ 9.6 Hz, H-2"), 5.25-5.07 (m, 2 H, CH_2 =), 5.02–4.98 (m, 2 H, H-4' H-3), 4.87 (d, 1 H, $J_{1'',2''}$ 8.0 Hz, H-1"), 4.83 (dd, 1 H, $J_{1',2'}$ 8.0, $J_{2',3'}$ 9.2 Hz, H-2'), 4.61 (d, 1 H, $J_{1',2'}$ 8.0 Hz, H-1'), 4.56–4.46 (m, 2 H), 4.43 (d, 1 H, $J_{1,2}$ 8.0 Hz, H-1), 4.24–4.20 (m, 2 H), 4.08-4.01 (m, 4 H), 3.85 (dd, 1 H, $J_{3',4'} = J_{2',3'} = 9.6$ Hz, H-3'), 3.64-3.59 (m, 2 H), 3.58 (dd, 1 H, $J_{1,2}$ 8.0, $J_{2,3}$ 9.6 Hz, H-2), 3.51-3.45 (m, 1 H), 3.38-3.29 (m, 1 H), 2.00, 1.98, 1.88, 1.86 (4 s, 12 H, 4 CH₃CO). Anal. Calcd for C₆₄H₆₄O₂₄: C, 63.15; H, 5.30. Found: C, 63.31; H, 5.17.

3.15. Allyl 2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 3)$ -[2,3,4,6-tetra-O-benzoyl- β -D-glucopyranosyl- $(1 \rightarrow 6)$]-2,4-di-O-acetyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ -4,6-O-benzylidene- α -D-glucopyranoside (23)

The 3-acetate of **23** (135 mg, 53% for two steps) was obtained as a syrup by coupling of **22** (235 mg, 0.15 mmol) with **5** (45 mg, 0.15 mmol), followed by acetylation under the same conditions as described for the preparation of the 3-acetate of **16**: $[\alpha]_D + 21.3^\circ$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.01–7.32 (m, 45 H, 9 Ph*H*), 5.88 (t, 1 H, *J* 9.6 Hz), 5.86 (t, 1 H, *J* 9.6 Hz), 5.82–5.75 (m, 1 H, =C*H*–), 5.68–5.6 (m, 2 H), 5.49 (s, 1 H, PhC*H*), 5.48–5.45 (m, 2 H), 5.37 (t, 1 H, *J* 9.6 Hz), 5.22–5.02 (m, 2 H), 5.01 (d, 1 H, *J* 8.0 Hz,

H-1), 4.88 (d, 1 H, J 3.6 Hz, H-1), 4.84 (d, 1 H, J 7.6 Hz, H-1), 4.79–4.70 (m, 2 H), 4.66–4.56 (m, 2 H), 4.49–4.44 (m, 2 H), 4.37 (d, 1 H, J 8.0 Hz, H-1), 4.18–4.09 (m, 3 H), 4.04–3.98 (m, 1 H), 3.94–3.88 (m, 2 H), 3.85–3.72 (m, 4 H), 3.68 (dd, 1 H, $J_{1,2}$ 2.8, $J_{2,3}$ 10.4 Hz, H-2), 3.62–3.54 (m, 2 H), 2.04, 1.90, 1.85 (3 s, 9 H, CH_3CO). Anal. Calcd for $C_{96}H_{88}O_{32}$: C, 65.75; H, 5.06. Found: C, 66.05; H, 4.98.

Acknowledgements

This work was supported by The Chinese Academy of Sciences (KZCX3-J-08) and by The National Natural Science Foundation of China (Projects 39970864 and 30070815).

References

- 1. (a); Khan, S. H.; O'Neil, R. A., Eds. *Modern Methods in Carbohydrate Synthesis*; Harwood Academic Publishers: United States, 1996; pp 125–150;
 - (b) Seeberger, P. H.; Eckhardt, M.; Utteridge, C. E.; Danishefsky, S. J. *J. Am. Chem. Soc.* **1997**, *119*, 10064–10070.
- (a) Wang, W.; Kong, F. J. Org. Chem. 1998, 63, 5744– 5745;

- (b) Wang, W.; Kong, F. Angew. Chem., Int. Ed. Engl. **1999**, 38, 1247–1250;
- (c) Zhu, Y.; Kong, F. Synlett 2001, 1217-1220;
- (d) Zhang, J.; Kong, F. *Tetrahedron: Asymmetry* **2002**, *13*, 243–252;
- (e) Ning, J.; Yi, Y.; Kong, F. Tetrahedron Lett. **2002**, 43, 5545–5549.
- 3. (a) Sasaki, T.; Takasuka, N. Carbohydr. Res. 1976, 47, 99-110;
 - (b) Kitamura, S.; Hori, T.; Kurita, K.; Takeo, K.; Hara, C.; Itoh, W.; Tabata, K.; Elgsaeter, A.; Stokke, B. T. *Carbohydr. Res.* **1994**. *263*. 111–120:
 - Carbohydr. Res. **1994**, 263, 111–120; (c) Chihara, G.; Maeda, Y.; Hamuro, J.; Sasaki, T.; Fukuoka, F. Nature **1969**, 222, 687–690;
 - (d) Schmid, F.; Stone, B. A.; McDougall, B. M.; Basic, A.; Martin, K. L.; Brownlee, R. T. C.; Chai, E.; Seviour, R. J. *Carbohydr. Res.* **2001**, *331*, 163–171.
- (a) Adeyeye, A.; Jansson, P. E.; Lindberg, B. Carbohydr. Res. 1988, 180, 295–299;
 - (b) Tokuyasu, K.; Ono, H.; Ohnishi-Kameyama, M.; Hayashi, K.; Mori, Y. *Carbohydr. Res.* **1997**, *303*, 453–458.
- (a) Muddasani, P. R.; Bernet, B.; Vasella, A. Helv. Chim. Acta 1994, 77, 257–290;
 - (b) Muddasani, P. R.; Bozo, E.; Bernet, B.; Vasella, A. Helv. Chim. Acta 1994, 77, 334-350.
- (a) Fraser-Reid, B.; Lopez, J. C.; Radhakrishnan, K. V.; Mach, M.; Schlueter, U.; Gomez, A. M.; Uriel, C. J. Am. Chem. Soc. 2002, 124, 3198–3199;
 - (b) Anilkumar, G.; Jia, Z. J.; Kraehmer, R.; Fraser-Reid, B. *J. Chem. Soc.*, *Perkin Trans.* 1 **1999**, 3591–3596;
 - (c) Anilkumar, G.; Nair, L. G.; Fraser-Reid, B. Org. Lett. **2000**, *2*, 2587–2589.